

CURRICULUM VITAE: Kevin H. Gardner

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OVERVIEW

Leading a multidisciplinary academic research group focused on determining how cellular machines sense and respond to environmental changes, gaining mechanistic insights into biology of oxygen and photosensing. Applied these discoveries to found two companies, leading to an FDA-approved targeted cancer therapy (belzutifan, Merck) and new optogenetic tools. Founded new interdisciplinary research center in urban public university, hired and mentoring 7 new faculty and established 3 core facilities. Advising strategic and technical facets of research, training, and entrepreneurship at international, national, city, and university levels.

ACADEMIC POSITIONS

Founding Director (2013-present)

Structural Biology Initiative
CUNY Advanced Science Research Center
City University of New York

Professor (2013-present)

Einstein Professor
Department of Chemistry and Biochemistry
City College of New York

Program Chair (2004-2010)

PI – NIH T32 GM008297 (2004-2014)

Graduate Program in Molecular Biophysics
Division of Basic Sciences, Southwestern Graduate
School of Biomedical Sciences
UT Southwestern Medical Center

Professor (2009-2013; adjunct 2013-2021)

Associate Professor (2004-2009)

Assistant Professor (1998-2004)

Departments of Biophysics, Biochemistry and Pharmacology
UT Southwestern Medical Center

TRAINING

Postdoctoral Research (1995-1998)

University of Toronto
Advisor: Lewis E. Kay, Ph.D.

Development of biophysical and biochemical methods to obtain structural models of larger proteins and protein complexes using solution NMR

Ph. D. (1989-1995)

Molecular Biophysics and Biochemistry
Yale University
Advisor: Joseph E. Coleman, M.D., Ph.D.

Cadmium-113 Heteronuclear NMR Studies of Zn₂Cys₆ DNA Binding Domains

B.S. (1985-1989, with Highest Honors):

Biochemistry, University of California, Davis

AWARDS AND HONORS

Stein & Moore Award

2023, Protein Society

BPS Biophysics of Health & Disease Award

2023, Biophysical Society

Keynote, Larock Undergraduate Research
Symposium

2017, UC Davis

Einstein Professor of Chemistry & Biochemistry

2013-present, City College of New York

Excellence in Education Awards

2010, 2012 UT Southwestern Medical Center

GRC Chairs' Hall of Fame

2011, Gordon Research Conferences

Excellence in Postdoctoral Mentoring Award

2011, UT Southwestern Postdoctoral Association

Virginia Lazenby O'Hara Chair in Biochemistry	2006-2013, UT Southwestern Medical Center
Outstanding Teacher Award	2004-2005, Dept. of Biochemistry, UT Southwestern
Meloche Lecturer	2004, Dept. of Chemistry, University of Wisconsin
CHS Hall of Fame	2000, Cupertino High School, Cupertino CA
Searle Scholar	1999-2002, The Chicago Community Trust
W.W. Caruth Jr. Scholar in Biomedical Research	1998-2013, UT Southwestern Medical Center
Helen Hay Whitney Postdoctoral Fellow	1996-1998, Helen Hay Whitney Foundation
NIH/NRSA Postdoctoral Fellow	1995-1996, National Institutes of Health
HHMI Predoctoral Fellow	1989-1994, HHMI
B.S. with Highest Honors (Biochemistry)	1989, UC Davis
Departmental Citation for Excellence	1989, UC Davis, Department of Biochemistry
Phi Beta Kappa	1989, UC Davis

INDUSTRIAL ACTIVITIES

2011-2019 Founding Consultant, Peloton Therapeutics, Inc., Dallas, TX

Peloton Therapeutics was a spinoff company developing small molecule HIF-2 α inhibitors initially discovered in my group in collaboration with Rick Bruick's lab, covered in Bruick et al. 2017 and Gardner et al. 2010 patents. I assisted with the spinoff, helping recruit funding and personnel at early stages, and maintained scientific advisory role throughout company life. Peloton successfully improved efficacy and suitability of nanomolar-potency academic lead compounds for clinical use, overseeing two HIF-2 α inhibitors through phase I, II clinical trials for clear cell renal cell carcinoma. Peloton raised over \$300M venture capital (led by The Column Group, with additional support by CPRIT) before May 2019 purchase by Merck for \$1.1B (with additional \$1.3B in incentives). Merck obtained FDA approval for HIF-2 α inhibitor WELIREG (belzutifan/MK-6482/PT-2977) clinical use in August 2021 in VHL-associated cancers and is currently in Phase III clinical trials as single agent and combination therapies for additional cancers.

2014-2018 Co-founder and CSO, Optologix, Inc., New York, NY & Dallas, TX

Optologix was a spinoff company from EL222 optogenetic light-regulated gene expression technology developed in my group, covered in Gardner et al. 2016 and 2019 patents. I developed the business plan within eLabNYC program with co-founder Laura Motta-Mena (2014-2015), jointly planned business and scientific strategy, and explored new product areas. Company operated 2015-2018, funded via sales of optogenetic tool kits, Health Wildcatters accelerator program (Dallas, TX), friends & family support.

PUBLICATIONS – PEER-REVIEWED RESEARCH AND REVIEW ARTICLES

96. Gagné, D., Azad, R., Edupuganti, U.R., Williams, J., Aramini, J.M., Akasaka, K. and Gardner, K.H. (2022) Use of high pressure NMR spectroscopy to rapidly identify proteins with internal ligand-binding voids. Manuscript under review; available at bioRxiv (DOI: 10.1101/2020.08.25.267195)

95. Dikiy, I., Swingle, D., Toy, K., Edupuganti, U.R., Rivera-Cancel, G. and Gardner, K.H. (2022) Conservation of function with diversification of higher-order structure within sensor histidine kinases. Manuscript under review; available at bioRxiv (DOI: 10.1101/2020.07.08.194225)

94. Berlew, E.E., Yamada, K., Rand, E.A., Kuznetsov, I.A., Ochs, C., Jaber, Z., Gardner, K.H. and Chow, B.Y. (2022) Designing single-component optogenetic membrane recruitment systems: The Rho-family GTPase signaling toolbox. *ACS Synthetic Biology*, **11**: 515-521. PMID: PMC8867532.

93. Hart, J.E. and Gardner, K.H. (2021) Lighting the way: Recent progress on the structure and function of phototropin blue light receptors. *J. Biol. Chem.*, **296**: 100594. PMID: PMC8086140.

92. Xu, X., Dikiy, I., Evans, M.R., Marcelino, L.P. and Gardner, K.H. (2021) Fragile protein folds: Sequence and environmental factors affecting the equilibrium of two interconverting, stably folded protein conformations. *Magn. Reson.*, **2**: 63-76. PMID: PMC9119131.

91. LaBelle, J., Ramos-Martinez, A., Shen, K., Motta-Mena, L.B., Gardner, K.H., Materna, S.C. and Woo, S. (2021) TAEL 2.0: An improved optogenetic expression system for zebrafish. *Zebrafish*, **18**: 20-28. PMID: PMC8020536.

90. Xu, X., Gagné, D., Aramini, J.M. and Gardner, K.H. (2021) Volume and compressibility differences between protein conformations revealed by high-pressure NMR. *Biophys. J.*, **120**: 924-935. (covered by Sprangers in *New and Notable* article in same issue, 749-751). PMID: PMC8008263.
89. Iuliano, J.N., Collado, J.T., Gil, A.A., Ravindran, P.T., Lukacs, A., Shin, S., Woroniecka, H.A., Adamczyk, K., Aramini, J.M., Edupuganti, U.R., Hall, C.R., Greetham, G.M., Sazanovich, I.V., Clark, I.P., Daryaei, T., Toettcher, J.E., French, J.B., Gardner, K.H., Simmerling, C.L., Meech, S.R. and Tonge, P.J. (2020) Unraveling the mechanism of a LOV domain optogenetic sensor: A glutamine lever induces unfolding of the α helix. *ACS Chem. Biol.*, **15**: 2752-2765. PMID: PMC7572778.
88. Dikiy, I. and Gardner, K.H. (2019) Shining light on the Alphaproteobacterial general stress response. *Mol. Micro.*, **112**: 438-441. PMID: PMC6703917.
87. Dikiy, I., Edupuganti, U.R., Abzalimov, R.R., Borbat, P.B., Srivastava, M., Freed, J.H. and Gardner, K.H. (2019) Insights into histidine kinase activation mechanisms from the monomeric blue light sensor EL346. *Proc. Natl. Acad. Sci.*, **116**: 4963-4972. PMID: PMC6421462.
86. Clark, L., Dikiy, I., Rosenbaum, D.M.[†] and Gardner, K.H.[†] (2018) On the use of *Pichia pastoris* for isotopic labeling of human GPCRs for NMR studies. *J. Biomol. NMR*, **71**: 203-211. ([†]: corresponding authors) PMID: PMC7282444.
85. Glantz, S.T., Berlew, E.E., Jaber, Z., Schuster, B.S., Gardner, K.H. and Chow, B.Y. (2018) Directly light-regulated binding of RGS-LOV photoreceptors to anionic membrane phospholipids. *Proc. Natl. Acad. Sci.*, **115**: E7720-E7727. PMID: PMC6099885.
84. Losi, A.[†], Gardner, K.H.[†] and Möglich, A.[†] (2018) Blue-light receptors for optogenetics. *Chem. Res.*, **118**: 10659-10709. ([†]: corresponding authors) PMID: PMC6500593.
83. Clark, L.^{*}, Dikiy, I.^{*}, Chapman, K., Rödström, K.E., Aramini, J., LeVine, M., Khelashvili, G., Rasmussen, S.G.F., Gardner, K.H.[†] and Rosenbaum, D.M.[†] (2017) Ligand modulation of sidechain dynamics in a wild-type human GPCR. *eLife*, **6**: e28505. PMID: PMC5650471. (^{*}:equal contributors, [†]: corresponding authors)
82. Reade, A., Motta-Mena, L.B., Gardner, K.H., Stanier, D.Y., Weiner, O.D. and Woo, S. (2017) TAE1: A zebrafish-optimized optogenetic gene expression system with fine spatial and temporal control. *Development*, **144**: 345-355. PMID: PMC5394756.
81. Corrêa, F., Key, J., Kuhlman, B. and Gardner, K.H. (2016) Computational repacking of HIF-2 α cavity replaces water-based stabilized core. *Structure*, **24**: 1918-1927.
80. Chen, W., Hill, H., Christie, A., Kim, M.S., Holloman, E., Pavía-Jiménez, A., Homayoun, F., Ma, Y., Patel, N., Yell, P., Hao, G., Yousuf, Q., Joyce, A., Pedrosa, I., Geiger, H., Zhang, H., Chang, J., Gardner, K.H., Bruick, R.K., Reeves, C., Hwang, T.H., Courtney, K., Frenkel, E., Sun, X., Zojwalla, N., Wong, T., Rizzi, J.P., Wallace, E.M., Josey, J.A., Xie, Y., Xie, X.-J., Kapur, P., McKay, R.M. and Brugarolas, J. (2016) Targeting renal cell carcinoma with a HIF-2 antagonist. *Nature*, **539**: 112-117. PMID: PMC5340502.
79. Corrêa, F. and Gardner, K.H. (2016) Basis of mutual domain inhibition in a bacterial signaling switch. *Cell Chem. Biol.*, **23**: 945-954. PMID: PMC5159254.
78. Glantz, S.T., Carpenter, E.J., Melkonian, M., Gardner, K.H., Boyden, E.S., Wong, G.K-S. and Chow, B.Y. (2016) Functional and topological diversity of LOV domain photoreceptors. *Proc. Natl. Acad. Sci. USA*, **113**: E1442-E1451. PMID: PMC4801262.
77. Scheuermann, T.H., Padrick, S.B., Gardner, K.H. and Brautigam, C.A. (2016) On the acquisition and analysis of microscale thermophoresis data. *Anal. Biochem.*, **496**: 79-93. PMID: PMC4873313.

76. Scheuermann, T.H., Stroud, D., Sleet, C., Bayeh, L., Shokri, C., Wang, H., Caldwell, C.G., Longgood, J., MacMillan, J.B., Bruick, R.K., Gardner, K.H. and Tambar, U.K. (2015) Isoform-selective and stereoselective inhibition of hypoxia inducible factor-2. *J. Med. Chem.*, **58**: 5930-5941.
75. Clark, L., Zahm, J.A., Ali, R., Kukula, M., Bian, L., Gardner, K.H., Rosen, M.K. and Rosenbaum, D.M. (2015) Methyl labeling and TROSY NMR spectroscopy of proteins expressed in the eukaryote *Pichia pastoris*. *J. Biomol. NMR*, **62**: 239-245. PMID: PMC4496254.
74. Guo, Y., Scheuermann, T.H., Partch, C.L., Tomchick, D.R. and Gardner, K.H. (2015) Coiled-coil coactivators play a structural role mediating interactions in hypoxia inducible factor heterodimerization. *J. Biol. Chem.*, **290**: 7707-7721. PMID: PMC4367273.
73. Ocasio, V., Corrêa, F. and Gardner, K.H. (2015) Ligand-induced folding of a two component signaling receiver domain. *Biochemistry*, **54**: 1353-1363. PMID: PMC4423417. (*Recommended in F1000 Prime; selected as featured article by journal*)
72. Rivera-Cancel, G., Ko, W.-H., Tomchick, D.R., Corrêa, F. and Gardner, K.H. (2014) Full-length structure of a monomeric histidine kinase reveals basis for sensory regulation. *Proc. Natl. Acad. Sci USA*, **111**: 17839-17844. PMID: PMC4273353. (*Recommended in F1000 Prime*)
71. Motta-Mena, L.B., Reade, A., Mallory, M.J., Glantz, S., Weiner, O.D., Lynch, K.W. and Gardner, K.H. (2014) An optogenetic gene expression system with rapid activation and deactivation kinetics. *Nat. Chem. Biol.*, **10**: 196-202. PMID: PMC3944926.
70. Salomon, D., Guo, Y., Kinch, L.N., Grishin, N.V., Gardner, K.H. and Orth, K. (2013) Effectors from animal and plant pathogens use a common domain to bind host phosphoinositides. *Nature Communications*, **4**: 2973.
69. Zoltowski, B.D., Motta-Mena, L.B. and Gardner, K.H. (2013) Blue-light induced dimerization of a bacterial LOV-HTH DNA-binding protein. *Biochemistry*, **52**: 6653-6661. PMID: PMC3813961.
68. Corrêa, F., Ko, W.-H., Ocasio, V., Bogomolni, R.A. and Gardner, K.H. (2013) Blue light regulated two-component systems: Enzymatic and functional analysis of light-oxygen-voltage (LOV)-histidine kinases and downstream response regulators. *Biochemistry*, **52**: 4656-4666. (*Selected as featured article by journal*) PMID: PMC3830641.
67. Freddolino, P.L., Gardner, K.H.[†] and Schulten, K.[†] (2013) Signaling mechanisms of LOV domains: New insights from molecular dynamics studies. *Photochem. Photobiol. Sci.*, **12**: 1158-1170 (†: *corresponding authors*) PMID: PMC3679247.
66. Rogers, J.L.*, Bayeh, L.*, Scheuermann, T.H.*, Longgood, J., Caldwell, C., Key, J., Naidoo, J., Melito, L., Shokri, C., Frantz, D.E., Bruick, R.K., Gardner, K.H.[†], MacMillan, J.B.[†] and Tambar, U.K.[†] (2013) Development of inhibitors of the PAS-B domain of the HIF-2 α transcription factor. *J. Med. Chem.*, **56**: 1739-1747. (*: *equal contributors*, †: *corresponding authors*) PMID: PMC3676484.
65. Moon, T.M., Corrêa, F., Kinch, L.N., Piali, A., Gardner, K.H.[†] and Goldsmith, E.J.[†] (2013) Solution structure of the WNK1 autoinhibitory domain, a WNK-specific PF2 domain. *J. Mol. Biol.*, **425**: 1245-1252. (†: *corresponding authors*)
64. Scheuermann, T.H., Li, Q., Ma, H.-W., Key, J., Zhang, L., Chen, R., Garcia, J.A., Naidoo, J., Longgood, J., Frantz, D.E., Tambar, U.K., Gardner, K.H.[†] and Bruick, R.K.[†] (2013) Allosteric inhibition of Hypoxia Inducible Factor 2 with small molecules. *Nat. Chem. Biol.*, **9**: 271-276. (†: *corresponding authors*) PMID: PMC3604136.

63. Guo, Y., Partch, C.L., Key, J., Card, P.B., Pashkov, V., Patel, A., Bruick, R.K., Wurdak, H. and Gardner, K.H. (2013) Regulating the ARNT-TACC3 axis: Multiple approaches to manipulating protein-protein interactions with small molecules. *ACS Chem. Biol.*, **8**: 626-635. PMID: PMC3600089.
62. Rivera-Cancel, G.* , Motta-Mena, L.B.* and Gardner, K.H. (2012) Identification of natural and artificial DNA substrates for the light-activated LOV-HTH transcription factor EL222. *Biochemistry*, **51**: 10024-10034. (*: equal contributors) PMID: PMC3531242.
61. Chollangi, S., Thompson, J.W., Ruiz, J.C., Gardner, K.H. and Bruick, R.K. (2012) The hemerythrin-like domain within F-box and leucine-rich repeat protein 5 communicates cellular iron and oxygen availability by distinct mechanisms. *J. Biol. Chem.*, **287**: 23710-23717. PMID: PMC3390645.
60. Zoltowski, B.D., Nash, A.I. and Gardner, K.H. (2011) Variations in protein/flavin hydrogen bonding in a LOV domain produce non-Arrhenius kinetics of adduct decay. *Biochemistry*, **50**: 8771-8779. (Accelerated Publication, selected as featured article by journal) PMID: PMC3381950.
59. Yuan, H., Dragnea, V., Wu, Q., Gardner, K.H. and Bauer, C.E. (2011) Mutational and structural studies of the PixD BLUF output signal that affects light-regulated interactions with PixE. *Biochemistry*, **50**, 6365-6375. PMID: PMC3139782.
58. Nash, A.I.* , McNulty, R.* , Shillito, M.E., Swartz, T.E., Bogomolni, R.A., Luecke, H.[†] and Gardner, K.H.[†] (2011) Structural basis of photosensitivity in a bacterial LOV-HTH DNA binding protein. *Proc. Natl. Acad. Sci.*, **108**: 9449-9454. (*: equal contributors, [†]: corresponding authors) PMID: PMC3111320.
57. Partch, C.L. and Gardner, K.H. (2011) Coactivators necessary for transcriptional output of the hypoxia inducible factor, HIF, are directly recruited by ARNT PAS-B. *Proc. Natl. Acad. Sci.*, **108**, 7739-7744. PMID: PMC3093465.
56. Song, S.-H., Freddolino, P.L., Nash, A.I., Carroll, E.C., Schulten, K., Gardner, K.H. and Larsen, D.S. (2011) Modulating LOV domain photodynamics with a residue alteration outside the chromophore binding site. *Biochemistry*, **50**, 2411-2423. (Accelerated Publication) PMID: PMC3068209.
55. Zoltowski, B.D. and Gardner, K.H. (2011) Tripping the light fantastic: Blue light photoreceptors as examples of environmentally-modulated protein:protein interactions. *Biochemistry*, **50**: 4-16. (selected as a featured article by journal) PMID: PMC3137735.
54. Akella, R., Min, X., Wu, Q., Gardner, K.H. and Goldsmith, E.J. (2010) The third stable conformation p38 α MAP kinase observed in phosphorylated p38 α and in solution. *Structure*, **18**: 1571-1578.
53. Strickland, D., Yao, X., Gawlak, G., Rosen, M.K., Gardner, K.H. and Sosnick, T.R. (2010) Rationally increasing the dynamic range of genetically-encoded, optically-controlled proteins. *Nature Methods*, **7**: 623-626. (covered in News and Views article, "Hold me tightly LOV" by K.M. Hahn and B. Kuhlman, *Nat. Methods* **7**(2010): 595-596; recommended in *F1000 Prime*) PMID: PMC2914111.
52. Lehotzky, R.E.* , Partch, C.L.* , Mukherjee, S., Cash, H., Goldman, W.E., Gardner, K.H.[†] and Hooper, L.V.[†] (2010) Molecular basis for peptidoglycan recognition by a bactericidal lectin. *Proc. Natl. Acad. Sci.*, **107**, 7722-7727. (*: equal contributors, [†]: corresponding authors; recommended in *F1000 Prime*) PMID: PMC2867859.
51. Key, J.* , Scheuermann, T.H.* , Anderson, P.C., Daggett, V. and Gardner, K.H. (2009) Principles of ligand binding within a completely buried cavity in HIF-2 α PAS-B. *J. Am. Chem. Soc.*, **131**: 17647-17654. (*: equal contributors) PMID: PMC2819816.
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49. Partch, C.L., Card, P.B., Amezcua, C.A. and Gardner, K.H. (2009) Molecular basis of coiled coil coactivator recruitment by the aryl hydrocarbon receptor nuclear translocator (ARNT) *J. Biol. Chem.*, **284**: 15184-15192. PMID: PMC2685699.
48. Wu, Q. and Gardner, K.H. (2009) BlrP1 BLUF domain: Structure and insight into light-induced changes. *Biochemistry*, **48**: 2620-2629. (Recommended in *F1000 Prime*)
47. Evans, M.R., Card, P.B. and Gardner, K.H. (2009) ARNT PAS-B has a fragile native state structure with an alternative beta-sheet register nearby in sequence space. *Proc. Natl. Acad. Sci.*, **106**: 2617-2622. PMID: PMC2650313.
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45. Scheuermann, T.H., Tomchick, D.R., Machius, M., Guo, Y., Bruick, R.K. and Gardner, K.H. (2009) Artificial ligand binding within the HIF-2 α PAS-B domain of the HIF2 transcription factor. *Proc. Natl. Acad. Sci.*, **106**: 450-455. PMID: PMC2626723.
44. Nash, A.I.*, Ko, W.-H.*, Harper, S.M. and Gardner, K.H. (2008) A conserved glutamine plays a central role in LOV domain signal transmission and duration. *Biochemistry*, **47**: 13842-13849. (*: equal contributors; selected as featured article by journal) PMID: PMC2630409.
43. Wu, Q., Ko, W.-H. and Gardner, K.H. (2008) Structural requirements for key residues and auxiliary portions of a BLUF domain. *Biochemistry*, **47**: 10271-10280.
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40. Lee, J., Tomchick, D.R., Brautigam, C.A., Machius, M., Kort, R., Hellingwerf, K.J. and Gardner, K.H. (2008) Changes at the KinA PAS-A dimerization interface influence histidine kinase function. *Biochemistry*, **47**: 4051-4064. (chosen as Hot Article by *Biochemistry*)
39. Kajimura, J., Rahman, A., Hsu, J., Evans, M.R., Gardner, K.H. and Rick, P.D. (2006) O-acetylation of the enterobacterial common antigen (ECA) polysaccharide is catalyzed by the product of the *yiaH* gene of *Escherichia coli* K-12. *J. Bacteriology*, **188**: 7542-7550. PMID: PMC1636290.
38. Colbert, C.L., Wu, Q., Erbel, P.J.A., Gardner, K.H. and Deisenhofer, J. (2006) Mechanism of substrate specificity in *Bacillus subtilis* ResA, a thioredoxin-like protein involved in cytochrome c maturation. *Proc. Natl. Acad. Sci.*, **103**: 4410-4415. PMID: PMC1400588.
37. Yang, J., Zhang, L., Erbel, P.J.A., Gardner, K.H., Ding, K.M., Garcia, J.A. and Bruick, R.K. (2005) Functions of the Per/ARNT/Sim (PAS) domains of the hypoxia inducible factor (HIF) *J. Biol. Chem.*, **280**: 36047-36054.
36. Card, P.B., Erbel, P.J.A. and Gardner, K.H. (2005) Structural basis of ARNT PAS-B dimerization: Use of a common β -sheet interface for hetero- and homodimerization. *J. Mol. Biol.*, **353**: 664-677.

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34. Jung, A., Domratcheva, T., Tarutina, M., Wu, Q., Ko, W.-H., Shoeman, R.L., Gomelsky, M., Gardner, K.H. and Schlichting, I. (2005) Structure of a bacterial BLUF photoreceptor: Insights into blue light-mediated signal transduction. *Proc. Natl. Acad. Sci. USA*, **102**: 12350-12355.
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28. Harper, S.M., Neil, L.C. and Gardner, K.H. (2003) Structural basis of a phototropin light switch. *Science*, **301**: 1541-1544. (*Recommended in F1000 Prime*)
27. Erbel, P.J.A., Barr, K., Chen, N., Gerwig, G.J., Rick, P.D. and Gardner, K.H. (2003) Identification and biosynthesis of cyclic ECA in *Escherichia coli*. *J. Bact.*, **185**: 1995-2004. PMID: PMC150143.
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25. Xiao, T., Gardner, K.H. and Sprang, S.R. (2002) Cosolvent-induced transformation of a death domain tertiary structure. *Proc. Natl. Acad. Sci.*, **99**: 11151-11156. PMID: PMC123225.
24. He, Q., Cheng, P., Yang, Y., Wang, L., Gardner, K.H. and Liu, Y. (2002) White collar-1, a DNA binding transcription factor as a light sensor. *Science*, **297**: 840-843. (*Recommended in F1000 Prime*)
23. Cheng, P., Yang, Y., Gardner, K.H. and Liu, Y. (2002) PAS domain-mediated WC-1/WC-2 interaction is essential for maintaining the steady-state level of WC-1 and the function of both proteins in circadian clock and light responses of *Neurospora*. *Mol. Cell. Biol.*, **22**: 517-524. PMID: PMC139750.
22. Holdeman, T.C. and Gardner, K.H. (2001) ^1H , ^{13}C and ^{15}N chemical shift assignments of the N-terminal PAS domain of mNPAS2. *J. Biomolecular NMR*, **21**: 383-384.
21. Usher, K.C., Özkan, E., Gardner, K.H. and Deisenhofer, J. (2001) The plug domain of FepA, a TonB-dependent transport protein from *E. coli*, binds its siderophore in the absence of the transmembrane barrel domain. *Proc. Natl Acad. Sci.*, **98**: 10676-10681. PMID: PMC58525
20. Rutter, J., Michnoff, C.H., Harper, S.M., Gardner, K.H. and McKnight, S.L. (2001) PAS kinase: an evolutionarily conserved PAS domain-regulated serine/threonine kinase. *Proc. Natl Acad. Sci.*, **98**: 8991-8996. PMID: PMC55361. PMID: PMC55361

19. Schweitzer, B.I., Foti, M., Keertikar, K., Kumar, S., Gardner, K.H. and Tucker-Kellogg, G. (1999) The use of ^{31}P relaxation experiments to probe the effects of nucleoside analogs on DNA dynamics. *Phosphorus, Sulfur and Silicon*, **144**: 301-304.

Postdoctoral research – University of Toronto (1995-1998)

18. Goto, N.K., Gardner, K.H., Mueller, G.A., Willis, R.C. and Kay, L.E. (1999) A robust and cost-effective method for the production of Val, Leu and Ile (δ^1) methyl-protonated ^{15}N , ^{13}C , ^2H -labeled proteins. *J. Biomol. NMR*, **13**: 369-374.

17. Gardner, K.H., Zhang, X., Gehring, K. and Kay, L.E. (1998) Solution NMR studies of a 42 kDa maltose binding protein/ β -cyclodextrin complex: chemical shift assignments and analysis. *J. Am. Chem. Soc.*, **120**: 11738-11748.

16. Zwahlen, C., Gardner, K.H., Sarma, S.P., Horita, D.A., Byrd, R.A. and Kay, L.E. (1998) An NMR experiment for measuring methyl-methyl NOEs in ^{13}C labeled proteins with high resolution. *J. Am. Chem. Soc.*, **120**: 7617-7625.

15. Yang, D., Gardner, K.H. and Kay, L.E. (1998) A sensitive pulse scheme for measuring the backbone dihedral angle ψ based on cross-correlation between $^{13}\text{C}\alpha$ - $^1\text{H}\alpha$ dipolar and carbonyl chemical shift anisotropy relaxation interactions. *J. Biomol. NMR*, **11**: 213-220.

14. Zwahlen, C., Vincent, S.J.F., Gardner, K.H. and Kay, L.E. (1998) Significantly improved resolution for NOE correlations from valine and isoleucine ($\text{C}\gamma^2$) methyl groups in ^{15}N , ^{13}C and ^{15}N , ^{13}C , ^2H -labeled proteins. *J. Am. Chem. Soc.*, **120**: 4825-4831.

13. Shan, X., Gardner, K.H., Muhandiram, D.R., Kay, L.E. and Arrowsmith, C.H. (1998) Subunit-specific backbone NMR assignments of a 64 kDa *trp* repressor/DNA complex: a role for N-terminal residues in tandem binding. *J. Biomol. NMR*, **11**: 307-318.

12. Gardner, K.H. and Kay, L.E. (1997) Production and incorporation of ^{15}N , ^{13}C , ^2H (^1H - δ^1 methyl) isoleucine into proteins for multidimensional NMR studies. *J. Am. Chem. Soc.*, **119**: 7599-7600.

11. Gardner, K.H., Rosen, M.K. and Kay, L.E. (1997) Global folds of highly deuterated, methyl protonated proteins by multidimensional NMR. *Biochemistry*, **36**: 1389-1401.

10. Gardner, K.H., Konrat, R., Rosen, M.K. and Kay, L.E. (1996) A (H)C(CO)NH-TOCSY pulse scheme for sequential assignment of protonated methyl groups in otherwise deuterated ^{15}N , ^{13}C labeled proteins. *J. Biomol. NMR*, **8**: 351-356.

9. Rosen, M.K., Gardner, K.H., Willis, R.C., Parris, W.E., Pawson, T. and Kay, L.E. (1996) Selective methyl group protonation of perdeuterated proteins. *J. Mol. Biol.*, **263**: 627-636.

8. Shan, X., Gardner, K.H., Muhandiram, D.R., Rao, N.S., Arrowsmith, C.H. and Kay, L.E. (1996) Assignment of the ^{15}N , $^{13}\text{C}\alpha$, $^{13}\text{C}\beta$ and HN resonances in an ^{15}N , ^{13}C , ^2H labeled 64 kDa *trp* repressor-operator complex using triple resonance NMR spectroscopy and ^2H -decoupling. *J. Am. Chem. Soc.*, **118**: 6570-6579.

Graduate research – Yale University (1989-1995) and Undergraduate research – UC Davis (1985-1989)

7. Gardner, K.H., Anderson, S.F. and Coleman, J.E. (1995) Solution structure of the *K. lactis* LAC9 Cd $_2$ Cys $_6$ DNA-binding domain. *Nat. Struct. Biol.*, **2**: 898-905.

6. Schweitzer, B.I., Gardner, K.H. and Tucker-Kellogg, G. (1995) HeteroTOCSY-based experiments for measuring heteronuclear relaxation in nucleic acids and proteins. *J. Biomol. NMR*, **6**: 180-188.

5. Klemba, M., Gardner, K.H., Marino, S., Clarke, N.D. and Regan, L. (1995) A novel family of designed metal-binding proteins. *Nat. Struct. Biol.*, **2**: 368-373 (addendum and correction: *NSB* **2**: 912)

4. Gardner, K.H. and Coleman, J.E. (1994) ^{113}Cd - ^1H heteroTOCSY: a method for determining metal-protein connectivities. *J. Biomol. NMR*, **4**: 761-774.
3. Schweitzer, B.I., Mikita, T., Kellogg, G.W., Gardner, K.H. and Beardsley, G.P. (1994) Solution structure of a DNA dodecamer containing the anti-neoplastic agent arabinosylcytosine: combined use of NMR, restrained molecular dynamics, and full relaxation matrix refinement. *Biochemistry*, **33**: 11460-11475.
2. Gardner, K.H., Pan, T., Narula, S., Rivera, E. and Coleman, J.E. (1991) Structure of the binuclear metal-binding site in the GAL4 transcription factor. *Biochemistry*, **30**: 11292-11302.
1. Sekiguchi, J.M., Cole, A.D., Gardner, K.H. and Kmiec, E.B. (1990) Transcription factor TFIIIA stimulates DNA supercoiling promoted by a fractionated cell-free extract from *Xenopus laevis*. *Eur. J. Biochem*, **192**: 311-320.

PUBLICATIONS – NON-PEER REVIEWED

16. Siclari, J.J. and Gardner, K.H. (2021) Two steps, one ligand: How PPAR γ binds small molecule agonists. *Structure*, **29**: 935-936. PMID: PMC8819882.
15. Dikiy, I., Clark, L., Gardner, K.H.[†] and Rosenbaum, D.M.[†] (2019) Isotopic labeling of eukaryotic membrane proteins for NMR studies of interactions and dynamics. *Meth. Enz.*, **614**: 37-65. ([†]: *corresponding authors*) PMID: PMC7309954.
14. Markley, J.L., Akutsu, H., Asakura, T., Baldus, M., Boelens, R., Bonvin, A., Kaptein, R., Bax, A., Bezsonova, I., Gryk, M.R., Hoch, J.C., Korzhnev, D.M., Maciejewski, M.W., Case, D., Chazin, W.J., Cross, T.A., Dames, S., Kessler, H., Lange, O., Madl, T., Reif, B., Sattler, M., Eliezer, D., Fersht, A., Forman-Kay, J., Kay, L.E., Fraser, J., Gross, J., Kortemme, T., Sali, A., Fujiwara, T., Gardner, K.H., Luo, X., Rizo-Rey, J., Rosen, M.K., Gil, R.R., Ho, C., Rule, G., Gronenborn, A.M., Ishima, R., Klein-Seetharaman, J., Tang, P., van der Wel, P., Xu, Y., Grzesiek, S., Hiller, S., Seeling, J., Laue, E.D., Mott, H., Nietlispach, D., Barsukov, I., Lian, L.Y., Middleton, D., Blumenschein, T., Moore, G., Campbell, I., Schnell, J., Vakonas, I.J., Watts, A., Conte, M.R., Mason, J., Pfuhl, M., Sanderson, M.R., Craven, J., Williamson, M., Dominguez, C., Roberts, G.C.K., Günther, U., Overduin, M., Werner, J., Williamson, P., Blindauer, C., Crump, M., Driscoll, P., Frenkiel, T., Golovanov, A., Matthews, S., Parkinson, J., Uhrin, D., Williams, M., Neuhaus, D., Oschkinat, H., Ramos, A., Shaw, D.E., Steinbeck, C., Vendruscolo, M., Vuister, G.W., Walters, K.J., Weinstein, H., Wüthrich, K. and Yokoyama, S. (2012) In support of the BMRB. *Nat. Struct. Mol. Biol.*, **19**: 854-860.
13. Rizo, J., Rosen, M.K. and Gardner, K.H. (2012) Enlightening molecular mechanisms through study of protein interactions. *J. Mol. Cell. Biol.*, **4**: 270-283. PMID: PMC3464395.
12. Gardner, K.H. and Corrêa, F. (2012) How plants see the invisible. *Science*, **335**: 1451-1452.
11. Gardner, K.H., Mittermaier, A. and Mulder, F.A.A. (2011) A tribute to Lewis Kay on his 50th birthday. *J. Biomol. NMR*, **51**: 3-4.
10. Motta-Mena, L.B., Partch, C.L. and Gardner, K.H. (2010) The three Rs of transcription: Recruit, retain and recycle. *Mol. Cell*, **40**: 855-858. PMID: PMC3012270.
9. Partch, C.L. and Gardner, K.H. (2010) Coactivator recruitment: A new role for PAS domains in transcriptional regulation by the bHLH-PAS family. *J. Cell. Physiol.*, **223**: 553-557. PMID: PMC2872778.
8. Gardner, K.H. (2008) Molecular Biophysics at UT Southwestern Medical Center: Strength through breadth. *Biopolymers*, **89**: 244-247.
7. Ko, W.-H., Nash, A.I. and Gardner, K.H. (2007) A LOVely view of blue light photosensing. *Nature Chem. Biol.*, **3**: 372-374.

6. Scheuermann, T.H., Yang, J., Zhang, L., Gardner, K.H. and Bruick, R.K. (2007) Hypoxia-Inducible Factor PAS domains: Structure and function. *Meth. Enz.*, **435**: 3-24.
5. Gardner, K.H. and Montminy, M. (2005) Can you hear me now: Regulation of transcriptional activators by phosphorylation. *Science STKE*, **2005**: pe 44.
4. Card, P.B. and Gardner, K.H. (2005) Identification and optimization of protein domains for NMR studies. *Meth. Enz.*, **394**: 3-16.
3. Gardner, K.H. and Kay, L.E. (1998) The use of ^2H , ^{13}C , ^{15}N multidimensional NMR to study the structure and dynamics of proteins. *Ann. Rev. Biophys. Biomol. Struct.*, **27**: 357-406.
(reprinted as Gardner, K.H. and Kay, L.E. (1999) Multidimensional ^2H -based NMR methods for resonance assignment, structure determination and the study of protein dynamics in Biological Magnetic Resonance: Modern Techniques in Protein NMR, **16**: 27-74)
2. Hardy, B.J., Doughty, S., Parretti, M., Tennison, J., Finn, B., Gardner, K.H. (1997) Internet conferences in nuclear magnetic resonance spectroscopy. *Prog. NMR Spec.*, **31**: 107-117.
1. Kay, L.E. and Gardner, K.H. (1997) Solution NMR spectroscopy beyond 25 kDa. *Curr. Op. Struct. Biol.*, **7**: 722-731.

PATENTS

6. Gardner, K.H., Motta-Mena, L.B. and Zoltowski, B.D. (2019) Blue-light inducible system for gene expression. US Patent Number 10,221,422 (issued March 5, 2019)
5. Bruick, R.K., Caldwell, C.J., Frantz, D.E., Gardner, K.H., MacMillan, J.B., Scheuermann, T.H. and Tambar, U.K. (2017) Inhibition of HIF-2 α heterodimerization with HIF-1 β (ARNT) US Patent Number 9,757,379 (issued September 12, 2017)
4. Gardner, K.H., Motta-Mena, L.B. and Zoltowski, B.D. (2016) Blue-light inducible system for gene expression. US Patent Number 9,506,073 (issued November 29, 2016)
3. Gardner, K.H., Amezcua, C.A., Erbel, P.J.A. and Card, P.B. (2010) NMR detection of foreign PAS domain ligands. US Patent Number 7,645,569 (issued January 12, 2010)
- 1,2. McKnight, S.L., Gardner, K.H., Harper, S., Rutter, J., Michnoff, C. and Amezcua, C. (2001) PAS kinase. US Patent Number 6,319,679 (issued November 20, 2001) and Patent Number 7,132,278 (issued November 7, 2006)

BOOK REVIEW

review of **Carbon-13 NMR Spectroscopy of Biological Systems** (Nicolau Beckmann, editor; 1995) *J. Magn. Reson. B* **111**(1996): 103-104.

RESEARCH FUNDING Track record of approx. \$40M raised in federal (NIH, NSF), state, foundation, and in-house funding as PI or co-I of proposals. Funded efforts include research in my group, collaborative multi-lab research, training, instrumentation, and infrastructure.

Current

- 2013-2025: NIH R01 GM106239: "Photosensory LOV proteins: Study and application of signaling mechanisms" (PI)
- 2018-2023: NSF MCB-1818148: "Protein Structural Defects: Sites for Small Molecule Binding and Regulation" (PI)
- 2021-2023: MSKCC-CCNY Partnership Pilot Grant: "Structural and Chemical Probing of A New Anticancer Target: HIF-Coactivator Complexes" (PI, with Derek Tan [MSKCC] co-I); awarded as part of U54 CA132378, "CCNY-MSKCC Partnership for Cancer Research, Education and Community Outreach"

- 2022-2023: CUNY Program Planning Grant: "A concerted experimental and computational attack to find small molecule inhibitors for 'undruggable' targets" (PI, with 5 co-Is developing NIH RM1 proposal)
- 2022-2024: NIH R21 GM147755: "A novel method to characterize cis-regulatory complexes during development" (co-I, with Mark Emerson [CCNY] PI)
- 2022-2025: Mathers Foundation MF-2112-02288: "Targeting HIF-associated factors Acss2 and CCCs: Essential and linked signal transducers in solid tumors" (PI, with Joseph Garcia [CUIMC] co-I)

Prior

- 2020-2021: NIH R01 GM106239-09S1: "AKTA PURE FPLC System for Macromolecular Purification and Characterization" (PI)
- 2019-2020: NIH R01 GM106239-08S1: "Helium Recovery System for CUNY ASRC/CCNY NMR Facilities" (PI)
- 2013-2017: CPRIT Individual Investigator Award RP130513, "Discovery and Optimization of Natural and Artificial Ligands Regulating Hypoxia Inducible Factor" (PI when awarded; subsequently switched to collaborator upon move from Texas)
- 1999-2014: Robert A. Welch Foundation I-1424: "Comparative studies of the signaling mechanisms of flavin-based protein photosensors" (PI)
- 2013-2016: CPRIT High Impact, High Risk RP130312: "Development of optogenetic tools for cellular and in vivo manipulation of cancer pathways" (PI when awarded; subsequently switched to collaborator upon move from Texas)
- 2012-2013: NIH S10 OD018027: "Acquisition of upgrades for 800MHz NMR console" (co-PI, with PI Jose Rizo-Rey)
- 2002-2013: NIH P01 CA095471: "A concerted chemical, biophysical and molecular biological attack on intracellular pathways relevant to cancer" (co-PI and directed 2 of 4 projects; PI: Steve McKnight)
- 2001-2012: NIH R01 GM081875: "Structural studies of PAS domain signaling mechanisms" (PI)
- 2009-2012: NIH R21 NS067624: "High Throughput Screen Development for Modulators of PAS/coactivator Interactions" (PI, with coinvestigator Rick Bruick)
- 2010: NIH R13 GM093350: "2010 Photosensory Receptors and Signal Transduction GRC/GRS Meetings" (PI)
- 2010-2013: CPRIT Individual Investigator Award RP100846, "Discovery and Optimization of Natural and Artificial Ligands Regulating Hypoxia Inducible Factor" (PI, with coinvestigators Rick Bruick, John MacMillan and Uttam Tambar)
- 2004-2014: NIH T32 GM008297: "Molecular Biophysics Predoctoral Training Program" (PI, renewed and transferred to Dr. Yuh Min Chook upon departure from UT Southwestern)
- 2004-2005: Advanced Technology Program (Texas Higher Education Coordinating Board) 010019-0124-2003: "Photoregulated enzymes: A novel and general approach to control function *in vitro* and *in vivo*" (PI, with coinvestigator Michael Rosen)
- 2002-2003: NIH S10 RR17270 (High End Instrumentation Grant): "800 MHz NMR spectrometer" (coinvestigator with PI: Jose Rizo-Rey)
- 1998-2002: Endowed Scholar in Biomedical Research Program, UT Southwestern Medical Center
- 1999-2002: Searle Scholar Program, Chicago Community Trust

TRAINEES *Proud of having mentored and mentoring approximately 60 trainees at postdoc, Ph.D., and undergraduate levels, with diversity of gender (ca. 1:1 women:men), racial and ethnic backgrounds, prior academic training, and career interests*

Postdoctoral researchers:

- Dr. Soumendu Boral (2023-present)
- Dr. Jaynee Hart (2019-2020) – *Postdoctoral Researcher, Robert Last Laboratory, Michigan State University*
- Dr. Meiling Zhang (2019-2021) – *Computational Biologist, Evozyne Inc., Chicago, IL*
- Dr. Igor Dikiy (NIH NRSA Postdoctoral Fellow 2015-2020; Finalist, Blavatnik Regional Young Scientist Award – Chemistry; NIH K22 awardee) – *Scientist – Protein Biochemistry, Regeneron Pharmaceuticals, Tarrytown, NY*
- Dr. Donald Gagné (Fonds de Recherche Québec Nature et Technologies Postdoctoral Fellow, 2015-2018) – *Research Scientist, Health Canada, Ottawa, Canada*
- Dr. Laura Motta-Mena (2010-2014; Senior Research Associate 2014-2016) – *Senior Associate, M Booth Health*

- Dr. Giomar Rivera-Cancel (2009-2014) – *Senior Research Associate, Deepak Nijhawan laboratory, UT Southwestern*
- Dr. Brian Zoltowski (NIH NRSA Postdoctoral Fellow 2008-2011) – *Associate Professor, Dept of Chemistry, Southern Methodist University*
- Dr. Fernando Corrêa (2008-2014; Senior Research Associate 2014-2016) – *Staff Scientist, Kodiak Sciences, Palo Alto, CA*
- Dr. Jason Key (2007-2011; Assistant Instructor 2011-2012) – *Associate Director of Technology and Innovation, SBGrid, Harvard Medical School*
- Dr. Carrie Partch (NIH NRSA Postdoctoral Fellow 2006-2011) – *Professor, Dept. of Chemistry and Biochemistry, UC Santa Cruz*
- Dr. Thomas Scheuermann (American Cancer Society Postdoctoral Fellow 2004-2010; Instructor 2010-2013) – *Senior Applications Engineer, Wyatt TechnologiesM*
- Dr. Qiong Wu (2003-2009) – *Director, Biomolecular NMR Facility, UT Southwestern*
- Dr. Paul Erbel (2000-2004) – *Research Investigator (Structural Sciences Unit), Novartis Institutes for Biomedical Research, Basel, CH*
- Dr. Carlos Amezcua (1999-2003) – *Associate Global R&D Fellow, FMC, Newark, DE*

Graduate students:

- Shaimaa Goher (2023-present; CUNY GC – Biochemistry Ph.D. Program)
- Malvin Forson (2023-present; CUNY GC – Biochemistry Ph.D. Program)
- Tarsisius Tiyani (2022-present; CUNY GC – Biochemistry Ph.D. Program)
- Joseph Closson (2022-present; CUNY GC – Biochemistry Ph.D. Program)
- James Siclari (2020-present; CUNY GC – Biology/MCD Ph.D. Program)
- Danielle Swingle (2019-present; CUNY GC – Biochemistry Ph.D. Program)
- Roksana Azad (2018-present; CUNY GC – Biochemistry Ph.D. Program; NIH F31 awardee)
- Matthew Cleere (2018-present; CUNY GC – Biology/MCD Ph.D. Program)
- Xingjian Xu (2018-present; CUNY GC – Biochemistry Ph.D. Program; 2022 Horst Schulz awardee for best first-authored paper by Biochemistry Ph.D. student)
- Dr. Uthama Edupuganti (2016-2021; CUNY GC – Biochemistry Ph.D. Program)
- Dr. Zaynab Jaber (2016-2021; CUNY GC – Biochemistry Ph.D. Program) – *Medical Associate, Health Science Communications, New York*
- Dr. Victor Ocasio-Ramirez (2010-2014; Ph.D. 2014) – *Field Applications Scientist, Nexcelcom, Lawrence, MA*
- Dr. Yirui Guo (2010-2014; Ph.D. 2014) – *Co-founder and CEO, Ligo Analytics (Dallas, TX) & MBA candidate, UT Dallas*
- Dr. Abigail Nash (MSTP, 2005-2009; completed Ph.D. portion of M.D./Ph.D. program 2009) – *Medical Director, Neurocrine Biosciences, San Diego, CA*
- Dr. Matthew Evans (2005-2009 - Ph.D. 2009; postdoc 2009-2010) – *Revenue Cycle Consultant, Acclaim Physician Group, Fort Worth, TX*
- Dr. Wen-Huang (Lisa) Ko (2004-2009; Ph.D. 2009) – *Head of Clinical Trial Monitoring, Dava Oncology, Dallas, TX*
- Dr. James Lee (MSTP, 2002-2006; completed Ph.D portion of M.D./Ph.D. program 2006) – *Pathologist in private practice, Alhambra, CA*
- Dr. Terra (Holdeman) Caudill (2000-2001) – *M.D. degree from UTMB; psychiatrist in private practice, Boca Raton, FL*
- Dr. Paul Card (2000-2005 - Ph.D. 2005; postdoc 2005-2006)
- Dr. Shannon Harper (2000-2004; Ph.D. 2004)
- (also mentored over 60 additional Ph.D. rotation students during first-year rotations in lab, 1999-present)

Undergraduate students: (both summer and academic-year work; home campus identified)

- Anastasiia Fisiuk (2023-present, CCNY – honors dissertation), Ramisha Aymon (2022-present, CCNY – honors dissertation), Jinho Seo (2022, CCNY), Keerthana Rameshbabu (2022, Texas A&M University), Faisal Younus (2021, CCNY), Divine Ehidom (2021, York College), Julia Gardner (2021, Duke University), Leandro Pimentel (2019-2022, CCNY – honors dissertation and postbac), Nichelle Camden (2019, Ausburg University), Kaitlyn Toy (2018-2021, CCNY/Macaulay – honors dissertation), Chyana Woodyard (2018, Hampton University), Nora Jaber (2018-2019, Hunter College); Megan Rodriguez (2018-2019, CCNY – honors dissertation); Mahmoud Soliman (2018, CCNY); Kenan Redzematovic (2018, CCNY); Sara

Wiener (2017, Yeshiva University); Katie Bunde (2017, CCNY); Casey Sniffin (2017-2018, Macaulay Honors College/CCNY); Andrew Palacios (2016-2018, Macaulay Honors College/CCNY and Columbia University); Aqib Muneer (2016, CCNY); Yun Young Kim (2016, Vassar College); Lucy Hovanisayan (2015, Univ. Pennsylvania); Kyle Ireton (2010, Oregon State Univ.); Laura Salguero (2008, New Mexico State Univ.); Leanna Steier (2007, Notre Dame); Aaron Maeng (2005, UT Austin); Daniel Buehler (2004, Univ. New Mexico); Brad Holmes (2001, Texas A&M Univ.); Mark Chan (1999, Harvard Univ.)

Sabbatical/Independent Scientist visitors:

Prof. Melanie Cocco (sabbatical visit, Fall 2023, UC Irvine)
 Dr. Carolina Matos (visiting postdoc, 2023-2024, Ramos & Almeida labs, UNICAMP, São Paulo, Brazil)
 Prof. Murray Whitelaw (sabbatical visit, Fall 2018, University of Adelaide)
 Dr. Arati Ramesh (independent postdoc, 2011-2014, Winkler lab, Univ. Maryland – College Park)
 Dr. Charles Dann (independent McKnight Fellow, 2005-2007, UT Southwestern)
 Dr. Susan Alguindigue (sabbatical visit, Summer 2002, Dept. of Chemistry, Univ. of Oklahoma)
 Prof. Ernest Blakeney (sabbatical visit, Fall 1999, Dept. of Chemistry, Centenary College)

PROFESSIONAL SERVICE – ADVISORY ROLES *Served and serving on advisory boards in four general areas: Structural biology operations at the international (RCSB PDB, iNEXT Discovery, Academia Sinica) and national scale (NYSBC, ACERT, NMRFAM), biotech entrepreneurship for New York City (LifeSciNYC: \$1B/10 yr initiative), advocacy panels for professional societies (Biophysical Society, ASBMB), academic community-building effort (CBSD)*

2023-2026	Member, Committee on Professional Opportunities for Women, Biophysical Society
2023-2025	Member, Institute of Biological Chemistry Academic Advisory Committee, Academia Sinica
2022-2025	Member, Public Affairs Action Committee, ASBMB
2022-present	Member, RCSB PDB Advisory Committee
2020-present	Member, SAB – iNEXT Discovery, European Union
2018-2023	Member, METRIC (Molecular Education, Technology and Research Innovation Center) External Advisory Board, NC State University
2017-present	Member, LifeSciNYC – Mayor’s Life Sciences Advisory Council, City of New York
2016-2021	Member, ACERT (National Biomedical Center for Advanced ESR Technology) External Advisory Board, Cornell University
2015-present	Member, Center for Biomolecular NMR Data Processing and Analysis BTTR External Advisory Board, UConn Health Science Center and University of Wisconsin
2014-present	Member, Board of Directors, NYSBC (New York Structural Biology Center)
2011-2016	Member, CBSD (Center for Biomolecular Structure and Dynamics) COBRE External Advisory Board, University of Montana
2009-2012	Member and Chair (2010-2012), NMRFAM (NMR Facility at Madison) External Advisory Board, University of Wisconsin, Madison

PROFESSIONAL SERVICE – CONFERENCE ORGANIZATION *Organized and organizing international and regional meetings in biochemistry, structural biology, chemical biology, and photosensing*

2022-2023	Co-organizer, Signaling Theme, 2023 DiscoverBMB – ASBMB Annual Meeting
2019-present	Member, Steering Committee, Chemical Biology Discussion Group, NY Academy of Sciences
2014-present	ICMRBS Council – Member (2014-present); Treasurer (2020-present)
2014	Chair, XXVI International Conference for Magnetic Resonance in Biological Systems (ICMRBS), Dallas, TX
2012	Organizer, Symposium on Biomolecular Structure, Dynamics and Function, Memphis, TN
2008, 2010	Vice-chair (2008) and Chair (2010) of Photosensory Receptors and Signal Transduction Gordon Research Conference

PROFESSIONAL SERVICE – GRANT, CENTER, AND MANUSCRIPT REVIEWS

2022	<i>Ad hoc</i> reviewer, NIEHS Structural Biology Laboratory
2021	<i>Ad hoc</i> member, NIH COBRE Review Study Sections (2) and RM1 Review Study Sections (2)

2019	Member, DFG SFB review panel
2018	Member, NSF MCB review panel
2017-2027	Member, Editorial Board, <i>Journal of Biological Chemistry</i>
2015-present	Member, Editorial Board, <i>Structure</i>
2015	Member, DFG SFB review panel
2014-2020	Permanent Member, NIH MSFC study section
2012	<i>Ad hoc</i> member, NIH MSFC study section
2012	<i>Ad hoc</i> reviewer, NCI Structural Biology Laboratory
2011-2013	Member, HHMI International Student Research Fellowship Review Committee
2011	<i>Ad hoc</i> member, NIH P01 Special Emphasis Panel
2009	<i>Ad hoc</i> member, NIH Stimulus panels, NIH/NCI Molecular Oncology P01 Special Emphasis Panel, NIH BBM Study Section
2008	<i>Ad hoc</i> member, NIH Shared Instrumentation Special Emphasis Panel and NIH MSFB Study Section
2006	<i>Ad hoc</i> member, NIH MSFB Study Section
2004	<i>Ad hoc</i> member, NIH BBCA Study Section
2001	<i>Ad hoc</i> member, NIH BBCA Special Study Section
ongoing	<i>Ad hoc</i> reviewer for <i>Science</i> , <i>Nature</i> , <i>Proc. Natl. Acad. Sci.</i> (editing and reviewing), <i>eLife</i> (editing and reviewing), <i>Biochemistry</i> , <i>Journal of the American Chemical Society</i> , <i>Journal of Molecular Biology</i> , and <i>Journal of Biomolecular NMR</i> and other journals. Also serving as an <i>ad hoc</i> reviewer for several funding agencies not noted above.

PROFESSIONAL SERVICE & TEACHING – CUNY

Service:

2022	CUNY System: Member, Strategic Planning Committee – Research & Innovation Subcommittee
2021	CUNY ASRC: Chair, Faculty Search Committee – ASRC Structural Biology Initiative; Member, Search Committee, Advancement Officer – Sciences
2015-2019	CUNY ASRC: Member, Biosafety Committee
2014-2017	CUNY ASRC: Chair, Faculty Search Committees (multiple) – ASRC Structural Biology Initiative
2021-present	CUNY Graduate Center: Representative, University Faculty Senate
2017-2020,	CUNY Graduate Center: Member, Executive Committee, Ph.D. Program in Biochemistry
2023-present	
2017-2020	CUNY Graduate Center: Member, Admissions and Awards Committee, Ph.D. Program in Biochemistry
2017-2020	CUNY Graduate Center: Member, Curriculum and Examination Committee, Ph.D. Program in Biochemistry
2015-present	CUNY Graduate Center: Faculty Member, Ph.D. Programs in Biochemistry, Chemistry and Biology
2015-present	Pre-tenure advisory committee member for CCNY, ASRC faculty (A. des Georges, S. Elbaum-Garfinkle, D. Keedy, R. Khayat, D. Eisele, S. Mingote, G. Grosso)
2015-present	Ph.D. dissertation committees (14 total in four graduate programs)

Teaching:

2016-2022	CCNY Chemistry 32002 – Biochemistry I; 30 hr lecture, 50-90 undergraduate students
2016	CCNY Chemistry 80541 – Molecular Biophysics Graduate Seminar; semester long, 8 Ph.D.
2015, 2018	GC Chemistry 86900 – Biomolecular NMR Spectroscopy; 3-6 hr lecture, 8 Ph.D. students
2015	GC Biochemistry 88800 – Hybrid Methods in Structural Biology; 3 hr lecture, 12 Ph.D. students

PROFESSIONAL SERVICE & TEACHING – UT SOUTHWESTERN

Service:

2013-2014	Department of Biophysics: Faculty Senate Representative
2012	Department of Biophysics: Faculty Search Committee

- 2002-2005 Department of Biochemistry: Computing Committee member
 1999-2012 Department of Biochemistry: Faculty Search Committee member (7 committees)
 1999-2012 Department of Biochemistry: Seminar and Events Committee member (chair: 1999-2002)
- 2004-2010 Molecular Biophysics: Graduate Program Chairman
 2004-2014 PI, NIH T32 GM008297: Molecular Biophysics Predoctoral Training Program
 2001-2005 Molecular Biophysics: Retreat Chairman
 2000-2014 Molecular Biophysics: Steering Committee member
 1999-2014 Ph.D. dissertation committees (38 total in five graduate programs)
 1999-2014 Ph.D. qualifying exam committees (22 total in two graduate programs)
- 2012-2014 member, Magnetic Resonance Safety Committee
 2010 Faculty Chair, UTSW Postdoctoral Association Symposium
 2007-2014 Inservice teaching and tour leader, STARS (Science Teacher Access to Resources at Southwestern) program
 2007-2010 member, search committee – Radiology Department chair

Teaching:

- 2010 DBS Core Course Biophysics Thread – co-director and 6 60' lectures, 15 Ph.D. topics: Biomolecular NMR Spectroscopy, Scattering Methods, Integration of Techniques
 2009 Biological Chemistry: Literature Discussion Group – 6 60' sessions, 10 Ph.D. students topics: Environmental sensing and signaling
 2008-2014 Molec. Biophysics: Advanced Biomolecular NMR – 4 90' lectures, 5 Ph.D./postdoc topics: NMR theory, pulse sequence development
 2008-2014 Molec. Biophysics: Modern Methods in Structural Biology – 4-6 90' lectures, 10 Ph.D. Course organizer and director topics: solution NMR – chemical shift assignment, structure determination
 2007-2013 Molec. Biophysics: Physical Biochemistry I – 1 90' lecture, 10-12 Ph.D. students topics: biophysical properties of proteins as polymers
 2003-2008 Molec. Biophysics: Enzymology / Physical Biochemistry II – 1-2 90' lectures, 4-10 Ph.D. topics: time-resolved biophysical methods; role of dynamics in catalysis
 2001, 2002 lecturer: Woods Hole Marine Biology Laboratory – Physiology course
 2001 lecturer: NRC/HHMI course “Determination of high-resolution structures for the post-genomic age”, Warsaw, Poland
 2001-2006 Molec. Biophysics: Biomolecular NMR – 3-5 90' lectures, 12 Ph.D. students topics: several topics in NMR theory and practice
 2000-2014 Medical Biochemistry – 5-6 60' lectures, 200-230 M.D. students topics: protein structure/function; allostery; hemoglobin
 1999-2007 DBS Core Course – 1-2 90' lectures, 60-95 Ph.D. students topics: protein NMR spectroscopy
 1999-2011 DBS Core Course – 5-6 90' literature review sessions, 8-10 Ph.D. students topics: protein structure and function

SOCIETY MEMBERSHIPS

American Chemical Society, Biophysical Society, AAAS, American Society for Biochemistry and Molecular Biology (ASBMB), New York Academy of Sciences

ADVOCACY AND PUBLIC OUTREACH ACTIVITIES *Participating in wide range of science outreach and discussion activities, ranging from Congressional visits to events at conferences and locally within NYC.*

- 2022-2025 Member, Public Advocacy Affairs Committee, ASBMB
 2023 Panelist, ASBMB Discover Careers Speed Networking session, DiscoverBMB 2023
 2023 Panelist, Yale Alumni Assoc. – Biotech Academia/Industry Panel in “Careers, Life, and Yale” series
 2022 Guest, Krea University’s “The Common Babblers” podcast, Sri City, India
 2022 Speaker, Taste of Science – AI/Drug Discovery/Structural Biology, New York, NY (virtual)

- 2022 Host, CUNY GC Presents “Demystifying Drug Discoveries” w/Michael Brown (UT Southwestern)
 2022 Participant, ASRC Visit, Sen. Chuck Schumer (NY)
 2022 Speaker, The Old Guard, West Hartford, CT
 2020 Participant, Virtual Rally for Biomedical Research, Washington, DC
 2020 Speaker, Secret Science Club, Brooklyn, NY
 2019 Guest, CUNY GC’s “The Thought Project – Episode 69” podcast, New York, NY
 2018 Capitol Hill Visit, Reps. Adriano Espillat (NY-13), Carolyn Maloney (NY-12), José Serrano (NY-15)
 2018 Speaker, Riverside Chat – Harlem Biospace, New York, NY
 2017 Host, ASRC Structural Biology Initiative Visit, Rep. Adriano Espillat (NY-13)
 2017 Host, CUNY GC Presents “Where Do Drugs Come From?” w/Neil Stahl (Regeneron), Neal Rosen (MSKCC), Laura Motta-Mena (Optologix)
 2015 Speaker, Science Exclamation Point, New York, NY

INVITED LECTURES (2020 – present)2023:

236. 2023 BPS Biophysics of Health & Disease Award Lecture, Biophysical Society (webinar)
 237. DiscoverBMB-ASBMB Conference, Seattle, WA
 238. Department of Chemistry & Biochemistry, University of Maryland, College Park, MD
 239. 19th Chemical Biophysics Symposium, University of Toronto, Toronto, Canada (Keynote Speaker)
 240. 37th Annual Meeting of the Protein Society, Boston, MA (Plenary Award Lecturer)
 241. Chemical Biology Symposium, New York University, NY
 242. Department of Chemistry, University of Nebraska, Lincoln, NE
 243. Eastern Analytical Symposium – EB Wilson Award Symposium for James Prestegard, Plainsboro, NJ

2022:

221. Brooklyn College Cancer Center “Diverse Approaches to Cancer Therapy” symposium, New York, NY
 222. Gordon Research Conference – Photosensory Receptors & Signal Transduction, Ventura, CA
 223. Department of Structural Biology, St. Jude Children’s Research Hospital, Memphis, TN
 224. NMR – A Tool for Biology, Pasteur Institute, Paris, France
 225. Department of Biochemistry & Molecular Biophysics, Columbia University Medical Center, New York, NY
 226. Department of Biochemistry, Weill Cornell Medical College, New York, NY
 227. XXIXth International Conference on Magnetic Resonance in Biological Systems, Boston, MA
 228. University College London – NMR Facility Dedication Symposium, London, UK
 229. Practical Applications of NMR in Industry Conference (PANIC), San Diego, CA
 230. Annual Symposium, NMR Topical Group, North Jersey ACS, Princeton, NJ
 231. Dartmouth Cancer Center, Hanover, NH
 232. Magnuson Center for Entrepreneurship, Dartmouth College, Hanover, NH
 233. New Equilibrium Biosciences, Cambridge, MA
 234. India|EMBO Lecture Course: Structure, Dynamics and Interactions in Biomolecular Systems Using NMR Spectroscopy, Berhampur, Odissa, India
 235. Krea University, Sri City, Andhra Pradesh, India

2021 (*: postponed or cancelled by COVID-19):

- . NMR – A Tool for Biology, Pasteur Institute, Paris, France*
 216. Cornell High Energy Synchrotron Source “Biology Under Extreme Conditions” meeting, Ithaca, NY (webinar)
 217. Canadian Chemistry Conference and Exhibition 2021, Winnipeg, MB (webinar)
 218. 20th International Symposium in Flavins and Flavoproteins, Graz, Austria
 219. Pitt/CMU Molecular Biophysics/Structural Biology Seminar Series, Pittsburgh, PA (webinar)
 220. Molecular & Cell Biology and Genetics Program, Drexel University School of Medicine, Philadelphia, PA (webinar)

2020 (*: postponed or cancelled by COVID-19):

212. Gordon Research Conference – Signal Transduction in Microorganisms, Ventura, CA
 213. Emerging Topics in Biomolecular Magnetic Resonance (webinar)
 214. Department of Biochemistry, University of Illinois – Champaign-Urbana, IL (webinar)
 215. SFB 1078 Protonation Dynamics in Protein Function colloquium (webinar)
 ---. Complex Carbohydrate Research Center, University of Georgia, Athens, GA*
 ---. Department of Chemistry, Berry College, Rome, GA*
 ---. Laboratory of Chemical Physics, NIH-NIDDK, Bethesda, MD*
 ---. Canadian Chemistry Conference and Exhibition 2020, Winnipeg, MB*
 ---. Gordon Research Conference – Photosensory Receptors & Signal Transduction, Ventura, CA*

- . 20th International Symposium in Flavins and Flavoproteins, Graz, Austria*
- . Center for Membrane Biology / Dept. of Biochemistry & Molecular Biology, UT Health, Houston, TX*
- . Pacificchem 2020, Honolulu, HI*